

3D time lapse imaging and quantitative analysis of the active migration of human vascular endothelial cells into a multilayered cell sheet with Yokogawa's confocal image cytometer CQ1

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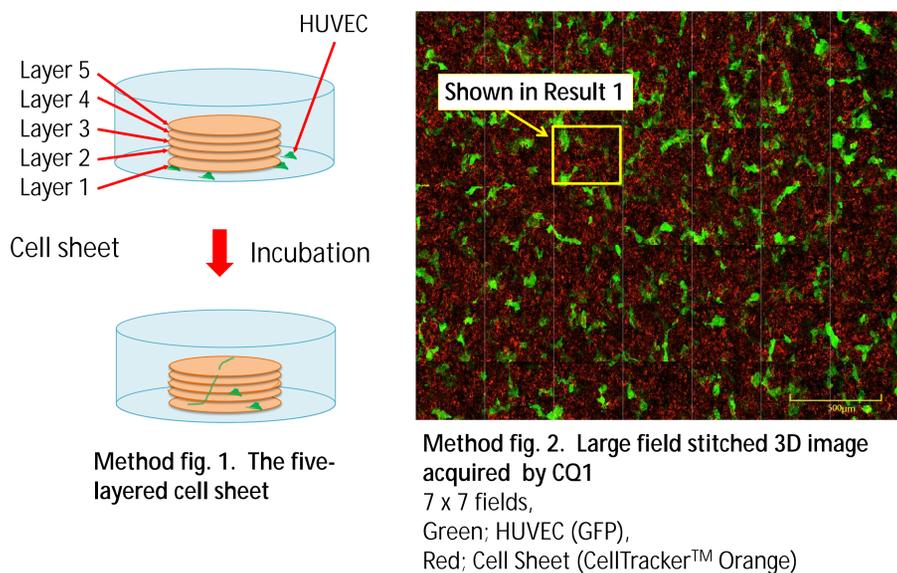
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Introduction

Time lapse confocal imaging has been an essential method to investigate the 3D dynamic behaviors of cells in tissue cultures. For long-term live cell imaging, it is critical to reduce phototoxic damage to the cells caused by repeated laser scanning. Yokogawa CSU (confocal scanner unit) is a confocal unit using a microlens-enhanced dual Nipkow disk confocal optical system, which has been shown to be less harmful to living cells compared to conventional single beam scanning devices. The CQ1 is an all-in-one confocal quantitative imaging cytometer based on the CSU. Here we report the 3D time lapse live cell imaging in a multilayered cell sheet using CQ1.

Methods

Five-layered myoblast cell sheets were constructed from human skeletal muscle myoblasts (HSM) and human skeletal muscle fibroblasts (HSMF) by using a thermo-responsive surface and stamp system⁽¹⁾⁽²⁾. HSMs and HSMFs were labeled with CellTracker™ Orange. Human umbilical vein endothelial cells (HUVEC) expressing GFP (GFP-HUVEC) were overlaid by the cell sheet and co-cultured. Time lapse imaging (67 hours, 30 min interval, 40x objective lens, 49 fields) was performed by CQ1 equipped with an internal incubation chamber to regulate culture environment.



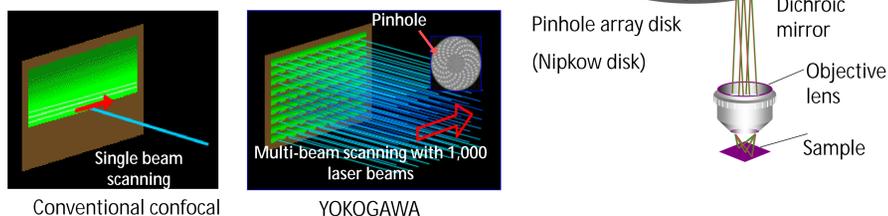
YOKOGAWA Core Competence

Microlens enhanced dual Nipkow disk scanning

→ high-speed, low photo toxicity and low photo bleaching

A Nipkow spinning disk and a second spinning disk with microlens rapidly scan the field of view with about 1,000 laser beams when rotated.

Multi-beam scanning not only increases scanning speed, but also results in significant lower photo toxicity and photo bleaching, because multiple excitation needs only a low level of laser power at the specimen to fully excite fluorescence.



Confocal Quantitative Image Cytometer CQ1

- Easy to use to measure the feature data of cells
- Cell clusters are directly measured by 3D imaging
- Gently 3D image acquisition
- Bench-top and no need for darkroom



Results

1. Dynamic migration and network formation of GFP-HUVECs captured by 3D time lapse imaging

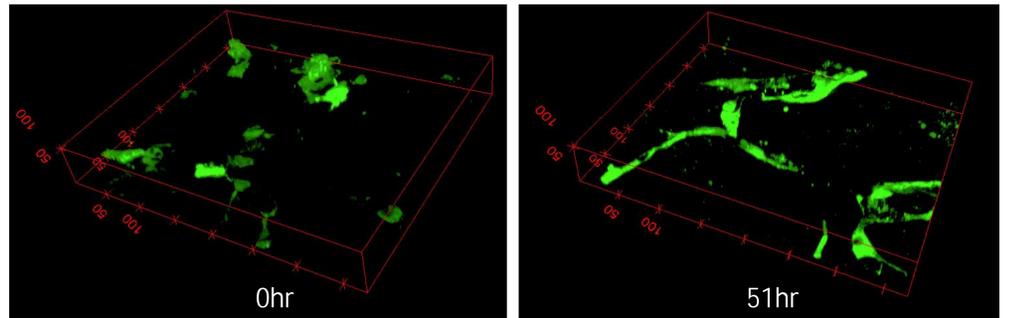


Fig. 1-1. 3D images of the cell sheet at the beginning of and after 51 hours incubation. Images were reconstructed from Z stack images of the field indicated by the yellow frame in the large field stitched image in Method fig.2.

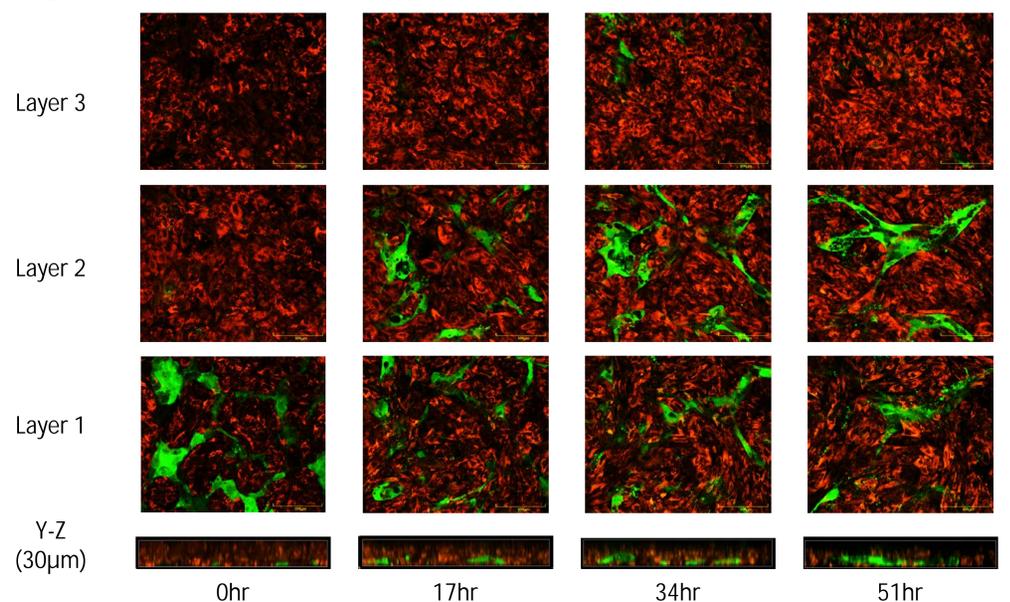


Fig. 1-2. Migration of the GFP-HUVECs into the cell sheet. (Rows, from top to bottom) Single slice images of layers 3, 2, 1 and corresponding Y-Z plane images of the cell sheet. (Columns, from left to right) Images acquired at 0, 17, 34 and 51 hr incubation. The image filed is the same as fig. 1-1.

2. Quantification of the migration of GFP-HUVECs into the five-layered cell sheet

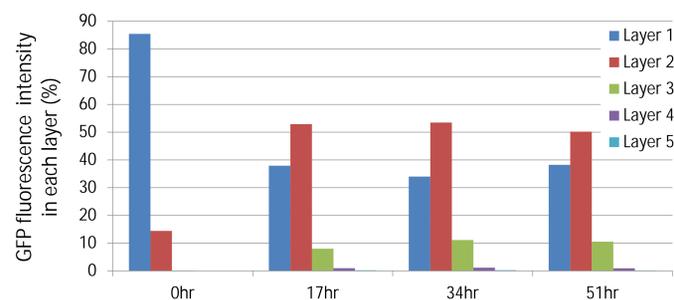


Fig. 2-1. Temporal change of the distribution GFP-HUVECs in the cell sheet. GFP fluorescence intensity in each layer was indicated as the ratio against the total GFP intensity in the cell sheet.

Summary & Conclusions

- Ø GFP-HUVECs dynamically migrated upward into the five-layered cell sheet constructed from HSMs and HSMFs.
- Ø The GFP-HUVECs formed a reticulate network in the horizontal plane in the middle layers.
- Ø Long-term 3D time lapse imaging by CQ1 revealed a dynamic process of the active migration and the formation of the cellular network in the multilayered cell sheet.
- Ø CQ1 would be a powerful research tool in tissue engineering as well as regenerative medicine and drug screening.

Acknowledgements

This study was conducted under the supervision of Dr. Nagamori, Osaka University and with Yokogawa Electric Corporation's responsibility.

Reference

- (1) Kino-oka et al., J. Biosci. Bioeng., 113, 128-131 (2012)
- (2) Nagamori et al., Biomaterials, 34, 662-668 (2013)